

# QBI Slide Scanners : Fluorescence Quick Guide

**Name:** .

**Group:** .

## Tissue Staining / Labelling:

## Pre-scan Classifier:

.prescan (5x) for 20x  
Integration times (camera gain 16):  
DAPI: .

## Scan Classifier:

20x  
Integration times (camera gain 5):  
DAPI: -  
488: -  
568: -  
647: -

## Scan Times:

Prescan: ~5 minutes  
Focus Map: ~5 minutes  
Final Scan (20x): ~25 minutes 1 colour  
2-4 hours for 3 colour

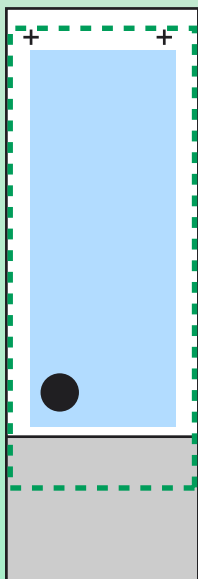
## Notes / Recommendations:

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## Instructions

1. Switch on the microscope and computer
2. Switch on the fluorescence Colibri LEDs
3. Load all of your slides - make sure they are **CLEAN, flat and pushed all the way to the top left in the slide frame**
4. Start Vslide - **START** online mode
5. Start the scanning software
6. Click **Adv. Mode**
7. Load an example slide and check its exposure settings:
  - i. Open the lamp shutter (foot pedal) and leave open
  - ii. Open the RL-shutter
  - iii. Select a fluorescent filter on the touch screen
  - iv. Right click on image window
  - v. Adjusting integration time with gain on 5 to get a good image without saturation
  - vi. Close the RL-shutter
8. Copy the integration times for each fluorescent channel into your **Scan classifier**
  - i. In the top menu: Metacyte>Classifier Setup
  - ii. Choose **your** Scan Classifier
  - iii. Select the **Capture** tab
  - iv. Change the integration times to those you observed or add channels if necessary
9. Click **Setup** and setup your slides:
  - i. Activate and Name each slide to be scanned (individually or in groups of 5)
  - ii. Choose your recommended **pre-scan classifier** under **Classifier**
  - iii. Under search window choose **Predefined area**
  - iv. Under size choose **Adv. Mode ++**
  - v. Ensure each activated slide has been named, and has the correct classifier selected as described above
  - vi. Click **OK**
10. Activate the robotic arm (right click **SF+** in the top right corner of the main screen and choose initialization)
11. Click **Scan**

## For Best Results



1. Use SuperFrost Plus slides
2. Use 24 x 60mm coverslips
3. Ensure some tissue / cells are positioned in the lower left corner (black dot)
4. Keep tissue away from the edges of the slide and below the ++ (in blue area)
5. If possible, use DAPI as a counter stain