

QBI Slide Scanners : Fluorescence Quick Guide

Name: .

Group: .

Tissue Staining / Labelling:

Pre-scan Classifier:

.prescan (5x) for 20x
Integration times (camera gain 16):
DAPI: .

Scan Classifier:

20x
Integration times (camera gain 5):
DAPI: -
488: -
568: -
647: -

Scan Times:

Prescan: ~5 minutes
Focus Map: ~5 minutes
Final Scan (20x): ~25 minutes 1 colour
2-4 hours for 3 colour

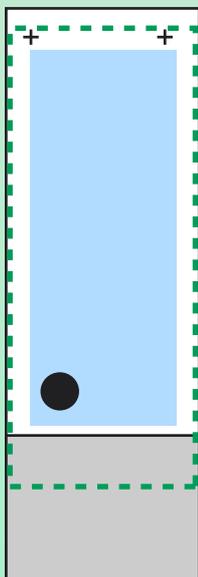
Notes / Recommendations:

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Instructions

1. Switch on the microscope and computer
2. Switch on the fluorescence Colibri LEDs
3. Load all of your slides - make sure they are **CLEAN, flat and pushed all the way to the top left in the slide frame**
4. Start Vslide - **START** online mode
5. Start the scanning software
6. Click **Adv. Mode**
7. Load an example slide and check its exposure settings:
 - i. Open the lamp shutter (foot pedal) and leave open
 - ii. Open the RL-shutter
 - iii. Select a fluorescent filter on the touch screen
 - iv. Right click on image window
 - v. Adjusting integration time with gain on 5 to get a good image without saturation
 - vi. Close the RL-shutter
8. Copy the integration times for each fluorescent channel into your **Scan classifier**
 - i. In the top menu: Metacyte>Classifier Setup
 - ii. Choose **your** Scan Classifier
 - iii. Select the **Capture** tab
 - iv. Change the integration times to those you observed or add channels if necessary
9. Click **Setup** and setup your slides:
 - i. Activate and Name each slide to be scanned (individually or in groups of 5)
 - ii. Choose your recommended **pre-scan classifier** under **Classifier**
 - iii. Under search window choose **Predefined area**
 - iv. Under size choose **Adv. Mode ++**
 - v. Ensure each activated slide has been named, and has the correct classifier selected as described above
 - vi. Click **OK**
10. Activate the robotic arm (right click **SF+** in the top right corner of the main screen and choose initialization)
11. Click **Scan**

For Best Results



1. Use SuperFrost Plus slides
2. Use 24 x 60mm coverslips
3. Ensure some tissue / cells are positioned in the lower left corner (black dot)
4. Keep tissue away from the edges of the slide and below the ++ (in blue area)
5. If possible, use DAPI as a counter stain